

# BIOSENSORS IN RAPID ANALYSIS FOR FOOD QUALITY

Ionela PRISACARI, Nina MIJA

Technical University of Moldova

**Abstract:** The need of manufacturers to efficiently control the production processes prompts the development of online and at line methods for quality control. Product quality cannot be sufficiently controlled only by recipes because it is not possible to check every single unit of a batch.

**Keywords:** biosensor and enzymatic reaction, on-line methods for analysis, biosensor for estimate of glucose content, biosensor for estimate monosodium glutamate acid content

The international standard of quality ISO 9000 was applied in 1990 and represents a point of transition from data quality control to whole systems of quality assurance.

In the aim of optimizing technology processes are created computer driven manufacturing sectors, which intercept sensor signals from sensors mounted in technological line. Then are accumulated a wide spread of data response is transmitted to setting mechanisms. Such technologies are called **sensory technologies** [8,9,12], and permit to provide measurements of process parameters automatically, in real time, which presents a significant opportunity in food production.

The area of using of sensors included: measure of moisture (40%), fat (23%), protein 14%, dry substances (6%), acidity (3%), other determination (14%)[1,7,10,12].

Recently, new type of sensors, biological sensors have been used (Fig.2). The difference from amperometric, electrochemical and other sensor is that biological sensor include same biological material (enzymes, antibodies, tissues in growth)

Construction of the biological sensor include 2 chief elements: a sensitive biological background, immobilized on a support, which may recognize a target substance despite of the presence of multicomponent environment. When the molecules are absorbed on the coated layer on the quartz crystal surface, the frequency of oscillation changes in the proportion to the amount of mass absorbed and the resulting change in current may be measured. The second element is a transducer, which firstly is capturing the changes in bioreceptors, then translate them into an electronic or an optical signal. A schematic presentation of an electronic sensor is shown in Fig. 1[7].

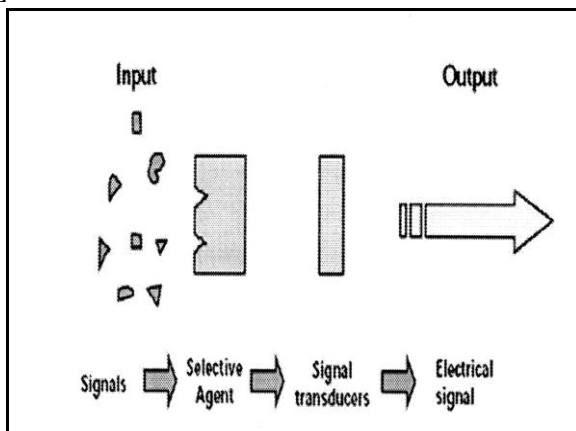


Fig. 1. Schematic presentation of electronic sensor

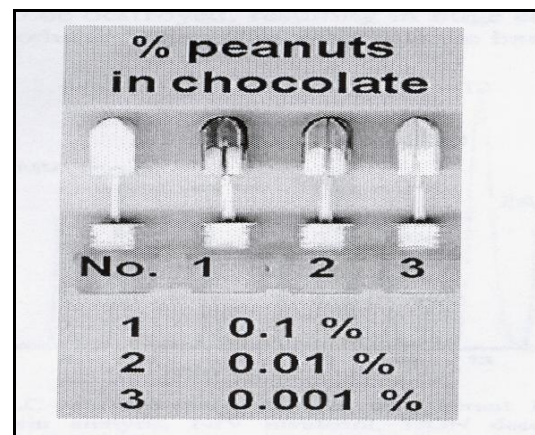
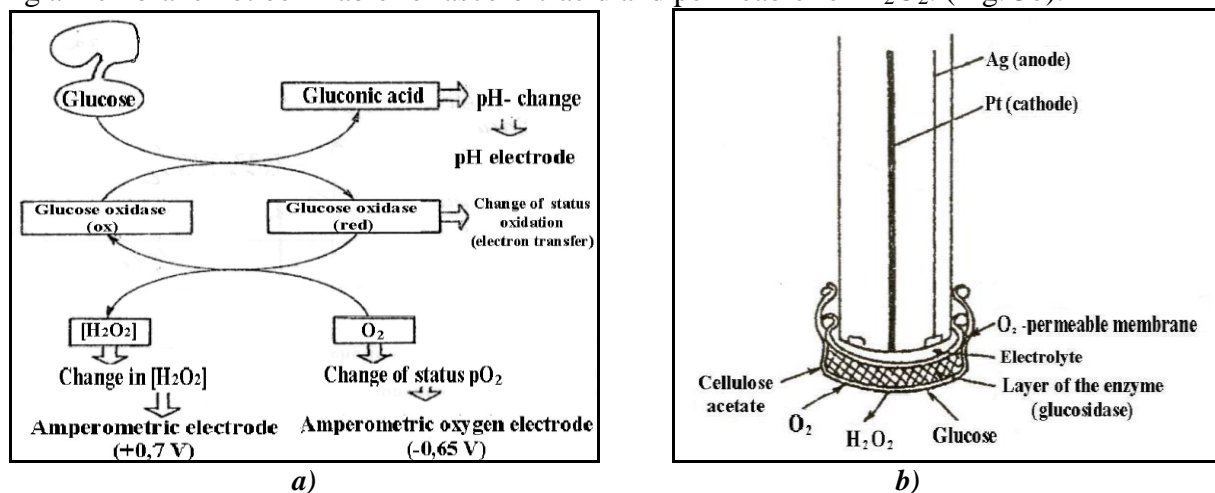


Fig. 2. Biosensor ELISA for determination of trace amounts of peanuts in protein extracts in a dip stick format suitable for production control.

**Sensor determination of glucose content** The first biosensor for estimate the glucose content was invented in 1962 by K.Clarc. It was for measure of electrochemical response in

changes in concentration of  $O_2/H_2O_2$ , the results or products of enzymatic reaction, which takes in the presence of enzyme glucozeoxidase (Fig.3a)[]. The advantage of this electrode consists in the working membrane that includes co-immobilized enzyme glucosoxidase. A layer of inert cellulose acetate will be used for separation of the prebe from enzymatic membrane.

The membranes of the electrode allowed the transition of  $H_2O_2$ , obtained as a final product of reaction enzyme & target substances. The second, external membrane is permeable for glucose and oxidase form of glucose gluconic acid. Electrode is polarized at +0,6 V & AgCl. Interference substances is ascorbic acid, which must oxidase on the Pt lay. The effect of ascorbic acid may be eliminated by using a membrane not permeable for ascorbic acid and permeable for  $H_2O_2$ . (Fig. 3b).



**Fig. 3.** Sensor determination of glucose: *a*) identification of measurable parameters for glucose determination based on biocatalytic oxidation reaction catalyzed by glucozeoxidase; *b*) Schematic construction of biosensor for glucose[4];

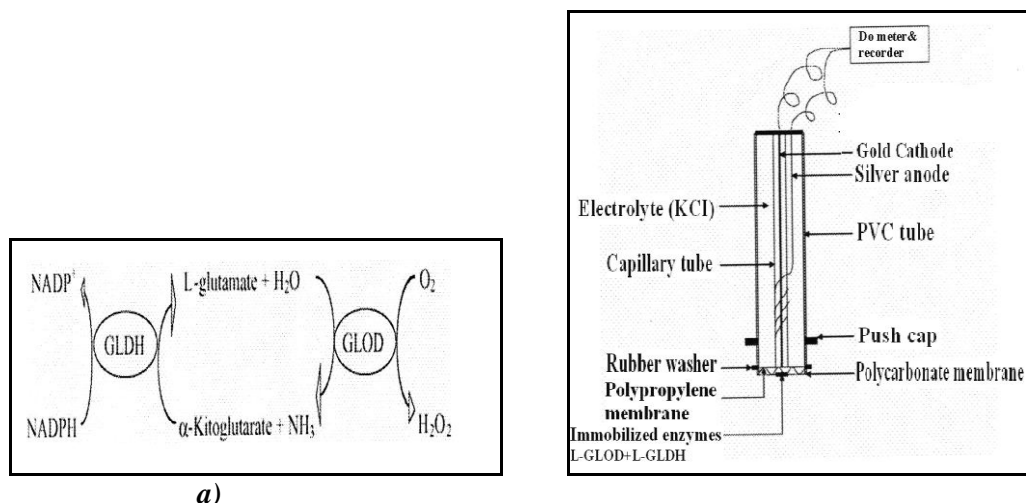
**Sensor determination of monosodium glutamate (MSG).** Amino acid L-glutamate is reported to be an important neuroexcitatory factor involved in several behavior patterns. Also it has widespread use as a flavor enhancing food additive. Thus, the determination of L-glutamate is of importance in both food and clinical samples. Glutamate concentration has been routinely assayed by chromatographic and potentiometric titration, but these methods are time consuming and require technical hand.

For the rapid estimation, several types of amperometric biosensor have been used. More recently, a biosensor made by co-immobilized L-glutamate oxidase (L-GLOD) and L-glutamate dehydrogenase (L-GLDH) as the bio-component that were incorporated into a modified electrode have been used for rapid estimation of monosodium glutamate (MSG).

Membranes of the electrode were prepared by binding in cross-linking enzymes on the polycarbonate substrate. In the presence of MSG, L-GLOD works as a cofactor and MSG is converted in kiloglutarate (Fig. 4a). Consumption of the  $O_2$  during the enzymatic reaction is registered by an oxygen recorder. Substrate recycling is continuous due to availability of 2 mM NADPH and 10 mM of ammonium ion in the system.

The electrode (1 cm in diameter) has 0,1M KCl solution as electrolyte, a silver anode and a gold cathode (with a sensitive end of 1,5 mm diameter). As shown in Fig. 4b the oxygen electrode was covered by one layer of enzymatic film on polycarbonate membrane (pore size 0,4  $\mu$ m) and was attached to the electrode by push cap system. Assays were performed in a 12 ml air-scaled experimental chamber, the solution being stirred constantly by a magnetic stirrer.

A. Basu, 2006 studied the value of MSG in real food samples (soy sauce, tomato sauce etc). This experimental data were compared with spectrophotometric data. A regression equation  $y=1,0642x-0,2289, R^2=0,998$  was obtained by the values of MSG found in food samples by spectrophotometric method versus sensor method.



**Fig. 4.** Determination of monosodium glutamate: **a)** the MSG converted to  $\alpha$ -kitoglutarate in enzymatic reaction; **b)** Schematic diagram of the MSG biosensor Assembly for  $O_2$  consumption (ppm/min)[2];

### Conclusion

1. Were created two generation of biosensors for determination of glucose content, but the apply is restricted *in vitro*, because biological materials included catalaza, an enzyme, which interference active aby redusing exces of  $H_2O_2$ .

2. A recycling electrode for determinatiuon of MSG may estimates probes with detection limits 0,02 mg MSG/L. The sensor had a half-life of over 60 days, with about 50 assays.

### References

1. Alocilja E.C., Radke S.M. *Market analysis of biosensors for food safety*// Biosensors & Bioelectronics, 2003, Vol.18, Issue 5-6, p. 841-847.
2. Basu A.K.A *biosensor based on co immobilized L-glutamate oxidase and L-glutamate dehydrogenase for analysis of monosodium glutamate in food*//Biosensors &Bioelectronics, 2006, 21 p. 1968-1972.
3. Beyene K., Negussie W. *A stable Glutamate Biosensor based on MnO >Bulk modified Screen-printed Carbon Electrode and Naflon film- immobilized Glutamate Oxidase* // African Journal of Chemistry, 2003, Vol. 56, p. 54-59.
4. Ciucanu I. *Metode fizico-chimice de analiză instrumentală*.Timișoara.Ed.Tehnică, 1986.-180p.
5. Gang C., Yating W. *Amperometric Biosensor coupled to Capillary electrophoresis for Glucose Determination* //Microchimica Acta, 2005, Vol. 150, Nr. 3-4. p. 239-245.
6. McMahon C.P., Rocchitta G. *Control of the Oxygen Dependence of an Implantable Polimer enzyme Composite Biosensor for Glutamate* //Analitical Chemistry, 2006, V-45, p. 453-458.
7. Muler A. Steihart H. *Recent developments in instrumental analysis for food quality*// Food Chemistry ,2007, Vol.102 , p.436-444.
8. Hansen T. *Sensory bazed qulity control utilizing an electronical nose and GC-MS analyses to predict end quality from raw materials* //Meat Sci., 2005, 69. Nr.4, p. 621-634.
9. Palleschi G., Cubadda R. *Electrochemical biosensors for food analysis and food industry*// Italian Journal of Food Science, 2001, Vol. 13, Issue 2., p. 137-158.
10. Sang H.L., Tai H.P. *Recent advances in the development of bioelectronic nose* // Biotechnology and Bioprocess Engineering, 2010, Vol. 15, Nr.1, p.22-29.
11. Yanling, G. *Quantification of water-soluble vitamins in milk-based infant formula using biosensor-based assays* //Food Chemistry,2008.Vol 110,Nr. 3 p.769-776.
12. Красников В.В. *Пищевая инженерия и качество продуктов*// Хранение и переработка сельхозсырья. 1996, № 2, с. 3-6.