

# Controlling the Risk of *Bacillus* in Food Using Berries

## Elisaveta Sandulachi, Viorica Bulgaru\*, Aliona Ghendov-Mosanu, Rodica Sturza\*

Technical University of Moldova, Chisinau, Republic of Moldova (RM) Email: \*rodica.sturza@chim.utm.md, \*viorica.bulgaru@tpa.utm.md

How to cite this paper: Sandulachi, E., Bulgaru, V., Ghendov-Mosanu, A. and Sturza, R. (2021) Controlling the Risk of *Bacillus* in Food Using Berries. *Food and Nutrition Sciences*, **12**, 557-577. https://doi.org/10.4236/fns.2021.126042

**Received:** May 24, 2021 **Accepted:** June 18, 2021 **Published:** June 21, 2021

Copyright © 2021 by author(s) and Scientific Research Publishing Inc. This work is licensed under the Creative Commons Attribution International License (CC BY 4.0). http://creativecommons.org/licenses/by/4.0/

## Abstract

Introduction: Increasing the shelf life of foods without the addition of synthetic additives is a demand from both producers and consumers. Spore-forming bacteria are a problem in the food industry. To reduce their impact, it is necessary to use complex technologies, as well as ingredients with antibacterial or antibiotic properties. The aim of this study was to develop initial symbiotic combinations between lactic acid bacteria and berries to control food quality. The relevant ability of lactic acid bacteria in the presence of berry additives to inhibit the growth of Bacillus strains that degrade bakery products and dairy products was investigated. The antibacterial effect of berries on the growth of Bacillus mesentericus was studied. Methods: In this study was used inhibition zone test, also called Kirby-Bauer Test. The growth rate of bacteria was based on the measurement of the optical density at 600 nm (OD600). The method of Thompson et al. has been used to research the development of ropiness disease in wheat bread samples. Results: The diameter of the Bacillus pumilus growth inhibition zones under the berries action was as follows: aronia  $-18.0 \pm 0.6$  mm; raspberry  $-16.0 \pm 0.4$  mm; strawberry  $-15.0 \pm 0.5$  mm. Lactic bacteria in the presence of berry additives showed a growth rate, measured by optical density (OD) at 600 from 0.073 to 0.651 (for aronia) from 0.071 to 0.609 (for raspberries), from 0.073 to 0.597 compared to the increase in environments without added fruit, which amounted to -0.050 to 0.410. In the yogurt with added fruit, a synergism was formed with Streptococcus thermophilus, Lactobacillus delbrueckii subsp. Bulgaricus, Lactococcus lactis subsp Lactis biovar diacetilactis. The influence of fat-soluble extracts of sea buckthorn, rose-hip, and hawthorn fruits on the development of ropiness disease when storing wheat bread was investigated. The general Pearson coefficient (microbial count and pH) for all fruit yogurt samples is 0.95066. Conclusion: The combined use of lactic acid bacteria and berries (aronia, raspberry and strawberry) had a synergistic effect on the risk posed by Bacillus bacteria. 1% of fat-soluble extract from the vegetable matter

reduced the risk of ropiness disease in wheat bread. This is due to the cumulative effect of the berries chemical composition (antioxidants, organic acids, etc.), increased acidity, lowered pH, and water activity of the food environment, below the development values of *Bacillus*.

#### **Keywords**

Risk of *Bacillus*, Berries, Lactic Acid Bacteria, Fermentation, Synergism, Bakery and Dairy Products

### **1. Introduction**

Preventing food spoilage without the addition of chemical food additives, while increasing functional properties of wheat-based bakery products, is an increasing demand by the consumers and a challenge for the food industry [1]. Functional food according to the European Commission Concerted Action on Functional Food Science in Europe (FuFoSE) and the International Life Sciences Institute (ILSI) is a food that beneficially affects one or more target functions in the body beyond adequate nutritional effects in a way that is relevant to either an improved state of health and well-being and/or reduction of a disease risk [2]. A food can be made functionally by increasing the concentration, adding, or improving the bioavailability of a particular component [3]. Certainly, the last decade, when consumer preferences have shifted to mildly processed food, new opportunities arise for spore-forming spoilage and pathogenic organisms [4].

Bakery products have a very short shelf life. Their quality depends on the time interval between baking and consumption [5]. Spoilage of bakery products is mainly due to the growth of moulds, the main species belonging to the genera *Aspergillus, Fusarium* and *Penicillium*, as well as to the roping of the bread, caused by *Bacillus sp.*, especially *Bacillus subtilis* and *Bacillus licheniformis* [6]. Statistics show that over the past decade, the number of small and medium-sized enterprises engaged in the production of bread and bakery products using non-traditional vegetable raw materials such as bran, seeds, grains, and legumes has increased. Such products are in demand among consumers, but the raw material is often a source of *Bacillus* bacteria, which requires the use of safe preservatives [7]. Considering the high microbiological contamination of raw materials (mainly flour) in the production of bread and bakery products, it is necessary to use complex technologies, as well as products with antimibacterial or antibiotic properties, to prevent the development of a ropiness disease and molding of bread [8].

Spore-forming bacteria can be a problem in the food industry, especially in the canning industry. Spores present in ingredients or present in the processing environment severely challenge the preservation process since their thermal resistance may be very high [9].

Cow's milk, which is one of the most complete foods [10], is at the same time

a favorable breeding ground for microorganisms [11] [12]. The quality and stability of dairy products are largely determined by the initial bacterial contamination and the composition of the raw material microflora [11] [12] [13]. Optionally anaerobic sporulating microorganisms (Bacillus subtillis, Bacillus mesentericus (Bacillus pumilus), Bacillus megatherium, Bacillus mycoides and Bacillus sereus) have pronounced proteolytic properties, coagulate, and peptonize milk, cause hemolysis, release ammonia, hydrogen sulfide [14]. Research in recent years has established that these are significant microorganisms of damage to dairy products and food poisoning [15] [16]. Recent studies conducted by us have shown that the risk control posed by bacteria of genus Bacillus could be achieved by using berries in food manufacturing recipes. Fruits, berries medicinal plants have bactericidal and bacteriostatic properties [17]-[23]. Therofere, Pashenko et al. [24] developed a technology for producing rowan leaven, which included rowan powder, beet sugar molasses and kefir fungus. Leaven is a water-molasses extract fermented with kefir fungus. The symbiosis of lactic acid bacteria and kefir yeast cells has fungicidal and bactericidal activity due to the high content of sorbic acid, tannins, phytoncides and flavonoids.

To ensure microbiological quality and consumer safety, strict controls and hygienic conditions are observed by the dairy industry, as recommended by guidelines of good dairy farming [25] [26] and hygienic conditions at dairy plants [27].

The control of environmental parameters (temperature, pH, water activity, salinity, atmosphere, presence of additives) can help the control of *Bacillus cereus* proliferation in foods. The ability to produce spores makes *Bacillus cereus* capable to escape processing conditions carried out by the food industries to preserve products and to eliminate or reduce the bacterial number in the final product [28]. Heat treatments commonly used by the food industry require longer treatments to efficiently eliminate *Bacillus cereus* spores [29].

The objective of the study was to evaluate the antimicrobial effect of different berries on the development of *Bacillus* bacteria in bakery and dairy products. Also, it was evaluated the combined effects of pasteurization intensity (no heat treatment and 10 min at 70°C, 80°C and 90°C), water activity ( $a_w$ ) (0.960 - 0.990), pH (5.5 - 7.0) and storage temperature (7°C and 10°C) on the survival and outgrowth of psychrotolerant spores of *Bacillus cereus* FF119b and *Bacillus pumilus* FF128a [30].

## References

- [1] Şimşek, Ö., Çon, A.H. and Tulumoğlu, Ş. (2006) Isolating Lactic Starter Cultures with Antimicrobial Activity for Sourdough Processes. *Food Control*, **17**, 263-270. <u>https://doi.org/10.1016/j.foodcont.2004.10.011</u>
- [2] Rositsa, D., Svetla, I., Zapryan, D., et al. (2014) Production of Wheat Bread without Preservatives Using Sourdough Starters. *Biotechnology & Biotechnological Equip*ment, 28, 889-898. <u>https://doi.org/10.1080/13102818.2014.965057</u>
- [3] Roberfroid, R. (2002) Functional Food Concept and Its Application to Prebiotics. *Digestive and Liver Disease*, 34, S105-S110. <u>https://doi.org/10.1016/S1590-8658(02)80176-1</u>

DOI: 10.4236/fns.2021.126042

571

Food and Nutrition Sciences

- [4] Abee, T., Groot, M.N., Tempelaars, M., Zwietering, M., et al. (2011) Germination and Outgrowth of Spores of *Bacillus cereus* Group Members: Diversity and Role of Germinant Receptors. *Food Microbiology*, 28, 199-208. https://doi.org/10.1016/j.fm.2010.03.015
- [5] Ghendov-Mosanu, A., Cristea, E., Patras, A., Sturza, R., *et al.* (2020) Potential Application of *Hippophae rhamnoides* in Wheat Bread Production. *Molecules*, 25, 1272. <u>https://doi.org/10.3390/molecules25061272</u>
- [6] Mentes, O., Ercan, R. and Akcelik, M. (2007) Inhibitor Activities of Two Lactobacillus Strains, Isolated from Sourdough, against Rope-Forming *Bacillus* Strains. *Food Control*, 18, 359-363. <u>https://doi.org/10.1016/j.foodcont.2005.10.020</u>
- [7] Alashbayeva, L., Shansharova, D., Mynbayeva, A., et al. (2021) Development of Technology for Bakery Products. Food Science and Technology (Campinas), 1-7. https://doi.org/10.1590/fst.61120
- [8] Rakhmonov, K.S., Atamuratova, T.I., Djuraeva, N.R., et al. (2020) Influence of Leavens of Spontaneous Fementation and Phytoadditives on the Provision of Microbiological Safety of Bread. Journal of Critical Reviews, 7, 850-860. https://doi.org/10.31838/jcr.07.05.177
- [9] Oomes, S.J., van Zuijlen, A.C., Hehenkamp, J.O., Witsenboer, H., van der Vossen, J.M. and Brul, S. (2007) The Characterisation of Bacillus Spores Occurring in the Manufacturing of (Low Acid) Canned Products. *International Journal of Food Microbiology*, **120**, 85-94. <u>https://doi.org/10.1016/j.ijfoodmicro.2007.06.013</u>
- [10] Sandulachi, E. and Bulgaru, V. (2019). Factors Affecting Quality of Goat's Milk Yogurt. Advances in Social Sciences Research Journal, 6, 205-221. <u>https://doi.org/10.14738/assrj.62.6129</u>
- [11] Nicolau, A. (2006) General Microbiology. Factors Influencing the Development of Microorganisms. Academica, Galati, 264. (In Romanian)
- [12] Sandulachi, E., Rubţov, S., *et al.* (2017) Microbiological Control of Food. Ed. Tehnica, TUM, 128 p. (In Romanian)
- [13] Dufrenne, J., Soentoro, P., Tatini, S., Day, T. and Notermans, S. (1994) Characteristics of *Bacillus cereus* Related to Safe Food Production. *International Journal of Food Microbiology*, 23, 99-109. <u>https://doi.org/10.1016/0168-1605(94)90225-9</u>
- [14] Koroleva, N.S. (1975) Technical Microbiology of Whole Milk Products. Food Industry, M, 271. (In Russian)
- [15] Borisova, G.V. (2001) Heat-Resistant Microorganisms Causing Defects in the Consistency and Organoleptic Properties of Dairy Products. Scientific Works of the All-Union State Agricultural Academy, 82-83. (In Russian)
- [16] Vasiliev, D.A., Zolotukhin, S.N., Feoktistova, N.A., *et al.* (2013) Biosensor Detection of Bacteria of the Genus Bacillus in Milk and Dairy Products to Prevent Spoilage. *Bulletin of the Ulyanovsk State Agricultural Academy*, **4**, 36-43. (In Russian)
- [17] Nile, S.H. and Park, S.W. (2014). Edible Berries: Bioactive Components and Their Effect on Human Health. *Journal of Nutrition*, **30**, 134-144. <u>https://doi.org/10.1016/j.nut.2013.04.007</u>
- [18] Sandulachi, E. (2018) Redox Properties of Strawberries and Raspberries. Lambert Academic Publishing, SIA Omni Scriptum Publishing, Latvia, 109. (In Russian)
- [19] Ghendov-Moşanu, A. (2018) Biologically Active Compounds of Horticultural Origin for Functional Foods. Ed. Tehnica-TUM, Chisinau, 236. (In Romanian)
- [20] Sandulachi, E., Cojocari, D., Balan, G., Popescu, L., Ghendov-Moşanu, A. and Sturza, R. (2020) Antimicrobial Effects of Berries on *Listeria monocytogenes. Journal of*

Food and Nutrition Sciences, 11, 873-886. https://doi.org/10.4236/fns.2020.119061

- [21] Sturza, R., Sandulachi, E., Cojocari, D., Balan, G., Popescu, L. and Ghendov-Moşanu, A. (2019) Antimicrobial Properties of Berry Powders in Cream Cheese. *Journal of En*gineering Science, **3**, 125-136.
- [22] Khan, M.F., Tang, H., Lyles, J.T., Pineau, R., Mashwani, Z.R. and Quave, C.L. (2018) Antibacterial Properties of Medicinal Plants from Pakistan against Multidrug-Resistant ESKAPE Pathogens. *Frontier in Pharmacology*, 9, 815. <u>https://doi.org/10.3389/fphar.2018.00815</u>
- [23] Munazir, M., Qureshi, R., Arshad, M. and Gulfraz, M. (2012) Antibacterial Activity of Root and Fruit Extracts of Leptadenia Pyrotechnica (Asclepiadaceae) from Pakistan. *Pakistan Journal of Botany*, **44**, 1209-1213.
- [24] Pashchenko, L.P., Kolomnikova, Ya.P., Ausheva, T.A. and Pashchenko, V.L. (2012) Biotechnological Aspects in Ensuring Microbiological Purity of Wheat Bread. *Voronezh State University of Engineering Technologies Bulletin*, 1, 87-89.
- [25] D.G 158 from 07-03-2019, Regarding the Approval of the Quality Requirements for Milk and Dairy Products. Official Gazette No. 111-118 Art. 218, Annex 4. (In Romanian)
- [26] IDF, F.A. (2011) Guide to Good Dairy Farming Practice. In Animal Production and Health Guidelines. International Dairy Federation and the Food and Agriculture Organization of the United Nations, Rome, 8.
- [27] ISO 8086:2004 [IDF 121:2004] Dairy Plant—Hygiene Conditions—General Guidance on Inspection and Sampling Procedures.
- [28] Vidic, J., Chaix, C., Manzano, M. and Heyndrickx, M. (2020) Food Sensing: Detection of *Bacillus cereus* Spores in Dairy Products. *Biosensors*, 10, 15. <u>https://doi.org/10.3390/bios10030015</u>
- [29] Dufrenne, J., Soentoro, P., Tatini, S., Day, T. and Notermans, S. (1994) Characteristics of *Bacillus cereus* Related to Safe Food Production. *International Journal of Food Microbiology*, 23, 99-109. <u>https://doi.org/10.1016/0168-1605(94)90225-9</u>
- [30] Samapundo, S., Heyndrickx, M., Xhaferi, R., de Baenst, I. and Devlieghere, F. (2014) The Combined Effect of Pasteurization Intensity, Water Activity, pH and Incubation Temperature on the Survival and Outgrowth of Spores of *Bacillus cereus* and *Bacillus pumilus* in Artificial Media and Food Products. *International Journal of Food Microbiology*, **181**, 10-18. https://doi.org/10.1016/j.ijfoodmicro.2014.04.018
- [31] Jessberger, N., Dietrich, R., Granum, P. and Martlbauer, E. (2020) The *Bacillus cereus* Food Infection as Multifactorial Process. *Toxins*, **12**, 701. https://doi.org/10.3390/toxins12110701
- [32] Abee, T., Groot, M.N., Tempelaars, M., Zwietering, M., Moezelaar, R. and van der Voort, M. (2011) Germination and Outgrowth of Spores of *Bacillus cereus* Group Members: Diversity and Role of Germinant Receptors. *Food Microbiology*, 28, 199-208. <u>https://doi.org/10.1016/j.fm.2010.03.015</u>
- [33] Jensen, G.B., Hansen, B.M., Eilenberg, J. and Mahillon, J. (2003) The Hidden Lifestyles of *Bacillus cereus* and Relatives. *Environmental Microbiology*, 5, 631-640. <u>https://doi.org/10.1046/j.1462-2920.2003.00461.x</u>
- [34] Vilain, S., Luo, Y., Hildreth, M.B. and Brozel, V.S. (2006) Analysis of the Life Cycle of the Soil Saprophyte *Bacillus cereus* in Liquid Soil Extract and in Soil. *Applied and Environmental Microbiology*, 72, 4970-4977. https://doi.org/10.1128/AEM.03076-05
- [35] Logan, N.A. and De Vos, P. (2015) Bacillus. In: Whitman, W.B., Ed., Bergey's Ma-

*nual of Systematics of Archaea and Bacteria*, John Wiley & Sons, Inc., in Association with Bergey's Manual Trust, Hoboken, 1-164. <u>https://doi.org/10.1002/9781118960608.gbm00530</u>

- [36] Stanish, L.F., Hull, N.M., Robertson, C.E., Harris, J.K., Stevens, M.J., Spear, J.R. and Pace, N.R. (2016) Factors Influencing Bacterial Diversity and Community Composition in Municipal Drinking Waters in the Ohio River Basin, USA. *PLoS ONE*, **11**, e0157966. <u>https://doi.org/10.1371/journal.pone.0157966</u>
- [37] Isnawati and Trimulyono, G. (2018) Temperature Range and Degree of Acidity Growth of Isolate of Indigenous Bacteria on Fermented Feed "Fermege". *Journal of Physics: Conference Series*, 953, Article ID: 012209. https://doi.org/10.1088/1742-6596/953/1/012209
- [38] Drobniewski, F.A. (1993) Bacillus cereus and Related Species. Clinical Microbiology Reviews, 6, 324-338. <u>https://doi.org/10.1128/CMR.6.4.324</u>
- [39] Brožová, M., Kubizniaková, P. and Matoulková, D. (2018) Brewing Microbiology—Bacteria of the Genera Bacillus, Brevibacillus and Paenibacillus and Cultivation Methods for Their Detection—Part 1. *Kvasny Prumysl*, 64, 50-57. <u>https://doi.org/10.18832/kp201813</u>
- Saleh, S.M., Harris, R.F. and Allen, O.N. (1970) Fate of *Bacillus thuringiensis* in Soil: Effect of Soil pH and Organic Amendment. *Canadian Journal of Microbiology*, 16, 677-680. <u>https://doi.org/10.1139/m70-116</u>
- [41] Priest, F.G. (2013) Bacillus. Colin R. Harwood. Springer Science & Business Media, Berlin.
- [42] Hudzicki, J. (2016) Kirby-Bauer Disk Diffusion Susceptibility Test Protocol. American Society for Microbiology, Washington DC, 23.
- [43] Hall, B.G., Acar, H., Nandipati, A. and Barlow, M. (2014) Growth Rates Made Easy. Molecular Biology and Evolution, 31, 232-238. https://doi.org/10.1093/molbev/mst187
- [44] Thompson, J.M., Waites, W.M. and Dodd, C.E.R. (1998) Detection of Rope Spoilage in Bread Caused by Bacillus Species. *Journal of Applied Microbiology*, 85, 481-486. <u>https://doi.org/10.1046/j.1365-2672.1998.853512.x</u>
- [45] Schoeni, J.L. and Wong, A.C. (2005) *Bacillus cereus* Food Poisoning and Its Toxins. *Journal of Food Protection*, 68, 636-648. <u>https://doi.org/10.4315/0362-028X-68.3.636</u>
- [46] Branda, S.S., González-Pastor, J.E., Ben-Yehuda, S., Losick, R. and Kolter, R. (2001) Fruiting Body Formation in *Bacillus subtilis. Proceedings of the National Academy* of Sciences of the USA, 98, 11621-11626. <u>https://doi.org/10.1073/pnas.191384198</u>
- [47] Garbeva, P., van Veen, J. and van Elsas, J. (2003) Predominant Bacillus spp. in Agricultural Soil under Different Management Regimes Detected via PCR-DGGE. *Microbial Ecology*, 45, 302-316. <u>https://doi.org/10.1007/s00248-002-2034-8</u>
- [48] Kramer, J.M. and Gilbert, R.J. (1989) *Bacillus cereus* and Other Bacillus Species. In: Doyle, M.P., Ed., *Foodborne Bacterial Pathogens*, Marcel Dekker, Inc., New York, 21-50.
- [49] Posada, R.H. (2017) Soil Microbiology in Agricultural Research in Relation to Ancient Knowledge. *Agricultural Research & Technology*, 6, 43-45. <u>https://doi.org/10.19080/ARTOAJ.2017.06.555685</u>
- [50] Anderson, Borge, G.I., Skeie, M., Sorhaug, T., Langsrud, T. and Granum, P.E. (2001) Growth and Toxin Profiles of *Bacillus cereus* Isolated from Different Food Sources. *International Journal of Food Microbiology*, **69**, 237-246.

https://doi.org/10.1016/S0168-1605(01)00500-1

- [51] Neysens, P. and De Vuyst, L. (2005) Kinetics and Modelling of Sourdough Lactic Acid Bacteria. *Trends Food Science & Technology*, **16**, 95-103. <u>https://doi.org/10.1016/j.tifs.2004.02.016</u>
- [52] Corsetti, A., Gobbetti, M., De Marco, B., Balestrieri, F., Paoletti, F., Russi, L. and Rossi, J. (2000) Combined Effect of Sourdough Lactic Acid Bacteria and Additives on Bread Firmness and Staling. *Journal of Agricultural and Food Chemistry*, 48, 3044-3051. <u>https://doi.org/10.1021/jf990853e</u>
- [53] Crowley, P., Schober, T., Clarke, C. and Arendt, E. (2002) The Effect of Storage Time on Textural and Crumb Grain Characteristics of Sourdough Wheat Bread. *European Food Research and Technology*, **214**, 489-496. <u>https://doi.org/10.1007/s00217-002-0500-7</u>
- [54] Thiele, C., Grassl, S. and Gänzle, M. (2004) Gluten Hydrolysis and Depolymerization during Sourdough Fermentation. *Journal of Agricultural and Food Chemistry*, 52, 1307-1314. <u>https://doi.org/10.1021/jf034470z</u>
- [55] Ganzle, M. and Vogel, R. (2002) Contribution of Sourdough Lactobacilli, Yeast and Cereal Enzymes to the Generation of Amino Acids in Dough Relevant for Bread Flavour. *Cereal Chemistry*, **79**, 45-51. <u>https://doi.org/10.1094/CCHEM.2002.79.1.45</u>
- [56] Liljeberg, H., Lonner, C. and Bjorck, I. (1995) Sourdough Fermentation or Addition of Organic Acids or Corresponding Salts to Bread Improves Nutritional Properties of Starch in Healthy Humans. *Journal of Nutrition*, **125**, 1503-1511.
- [57] Corsetti, A., Gobbetti, M., Rossi, J. and Damiani, P. (1998) Antimould Activity of Sourdough Lactic Acid Bacteria: Identification of a Mixture of Organic Acids Produced by *Lactobacillus sanfrancisco* CB1. *Applied Microbiology and Biotechnology*, 50, 253-256. <u>https://doi.org/10.1007/s002530051285</u>
- [58] Malthew, A., Renschler, A.W., Nnandi, A., et al. (2020) Using Nitrous Acid-Modified de Man, Rogosa, and Sharpe Medium to Selectively Isolate and Culture Lactic Acid Bacteria from Dairy Foods. Journal of Dairy Science, 103, 1215-1222. https://doi.org/10.3168/jds.2019-17041
- [59] Marín, S., Abellana, M., Rubinat, M., Sanchis, V. and Ramos, A.J. (2003) Efficacy of Sorbates on the Control of the Growth of Eurotium Species in Bakery Products with near Neutral pH. *International Journal of Food Microbiology*, 87, 251-258. <u>https://doi.org/10.1016/S0168-1605(03)00068-0</u>
- [60] Guynot, M.E., Ramos, A.J., Sanchis, V. and Marín S. (2005) Study of Benzoate, Propionate, and Sorbate Salts as Mould Spoilage Inhibitors on Intermediate Moisture Bakery Products of Low pH (4.5-5.5). *International Journal of Food Microbiology*, 101, 161-168. https://doi.org/10.1016/j.ijfoodmicro.2004.11.003
- [61] Corsetti, A. and Settanni, L. (2007) Lactobacilli in Sourdough Fermentation. Food Research International, 40, 539-558. <u>https://doi.org/10.1016/j.foodres.2006.11.001</u>
- [62] Guynot, M., Marin, S., Sanchis, V. and Ramos, A. (2005) An Attempt to Optimize Potassium Sorbate Use to Preserve Low pH (4.5-5.5) Intermediate Moisture Bakery Products by Modelling Eurotium spp., Aspergillus spp. and *Penicillium corylophilum* Growth. *International Journal of Food Microbiology*, **101**, 169-177. https://doi.org/10.1016/j.ijfoodmicro.2004.11.002
- [63] Albiac, M.A., Di Cagno, R., Filannino, P., Cantatore, V. and Gobbetti, M. (2020) How Fructophilic Lactic Acid Bacteria May Reduce the FODMAPs Content in Wheat-Derived Baked Goods: A Proof of Concept. *Microbial Cell Factories*, 19, 182. https://doi.org/10.1186/s12934-020-01438-6

- [64] Zougagh, S., Belghiti, A., Rochd, T., Zerdani, I. and Mouslim, J. (2019) Medicinal and Aromatic Plants Used in Traditional Treatment of the Oral Pathology: The Ethnobotanical Survey in the Economic Capital Casablanca, Morocco (North Africa). *Natural Product and Bioprospecting*, 9, 35-48. https://doi.org/10.1007/s13659-018-0194-6
- [65] Black, E.P., Wei, J., Atluri, S., Cortezzo, D.E., Koziol-Dube, K., Hoover, D.G. and Setlow, P. (2007) Analysis of Factors Influencing the Rate of Germination of Spores of *Bacillus subtilis* by Very High Pressure. *Journal of Applied Microbiology*, **102**, 65-76. <u>https://doi.org/10.1111/j.1365-2672.2006.03062.x</u>
- [66] Ahmed, A.H., Moustafa, K. and Marth, E.H. (1983) Incidence of *Bacillus cereus* in Milk and Some Milk Products. *Journal of Food Protection*, 46, 126-128. <u>https://doi.org/10.4315/0362-028X-46.2.126</u>
- [67] Mikolajcik, E.M., Kearney, J.W. and Kristoffersen, T. (1973) Fate of *Bacillus cereus* in Cultured and Direct Acidified Skim Milk and Cheddar Cheese. *Journal of Milk* and Food Technology, 36, 317-320. <u>https://doi.org/10.4315/0022-2747-36.6.317</u>
- [68] Cortezzo, D.E., Koziol-Dube, K., Setlow, B. and Setlow, P. (2004) Treatment with Oxidizing Agents Damages the Inner Membrane of Spores of *Bacillus subtilis* and Sensitizes Spores to Subsequent Stress. *Journal of Applied Microbiology*, 97, 838-852. https://doi.org/10.1111/j.1365-2672.2004.02370.x
- [69] Saikia, D. and Sit, N. (2014) Effect of Using Sourdough and Frozen Dough for Preparation of Breads on Quality, Shelf Life and Staling. *American Journal of Food Technology*, 9, 223-230. <u>https://doi.org/10.3923/ajft.2014.223.230</u>
- [70] Sadeghi, A.F., Shahidi, S.A., Mortazavi and Sadeghi, B. (2008) Evaluation of Sourdough Effect on Microbiological Shelf Life and Sensory Properties of Iranian Barbari Bread. *Biotechnology*, 7, 354-356. <u>https://doi.org/10.3923/biotech.2008.354.356</u>
- [71] Dal Bello, F., Clarke, C.I., Ryan, L.A.M., Ulmer, H., Schober, T.J., et al. (2007) Improvement of the Quality and Shelf Life of Wheat Bread by Fermentation with the Antifungal Strain *Lactobacillus plantarum* FST 1.7. *Journal of Cereal Science*, 45, 309-318. <u>https://doi.org/10.1016/j.jcs.2006.09.004</u>
- [72] Strom, K., Sjogren, J., Broberg, A. and Schnurer, J. (2002) Lactobacillus plantarum MiLAB 393 Produces the Antifungal Cyclic Dipeptides Cyclo(l-Phe-l-Pro) and Cyclo(l-Phe-trans-4-OH-l-Pro) and 3-Phenyllactic Acid. Applied Environmental Microbiology, 68, 4322-4327. <u>https://doi.org/10.1128/AEM.68.9.4322-4327.2002</u>
- [73] Arena, M.P., Silvain, A., Normanno, G., Grieco, F., Drider, D., Spano, G. and Fiocco, D. (2016) Use of *Lactobacillus plantarum* Strains as a Bio-Control Strategy against Food-Borne Pathogenic Microorganisms. *Frontiers in Microbiology*, 7, 464. https://doi.org/10.3389/fmicb.2016.00464
- [74] Katina, K., Msauri, H., Alakomi, H.L. and Mattila-Sandholm, T. (2002) Potential of Lactic Acid Bacteria to Inhibit Rope Spoilage in Wheat Sourdough Bread. *LWT—Food Science and Technology*, 35, 38-45. <u>https://doi.org/10.1006/fstl.2001.0808</u>
- [75] Carlin, F. (2011) Origin of Bacterial Spores Contaminating Foods. Food Microbiology, 28, 177-182. <u>https://doi.org/10.1016/j.fm.2010.07.008</u>
- [76] Hornstra, L.M., Ter Beek, A., Smelt, J.P., Kallemeijn, W.W. and Stanley, B. (2009) On the Origin of Heterogeneity in (Preservation) Resistance of Bacillus Spores: Input for a "Systems" Analysis Approach of Bacterial Spore Outgrowth. *International Journal of Food Microbiology*, **134**, 9-15. https://doi.org/10.1016/j.ijfoodmicro.2009.02.011
- [77] Andriani, Y., Safitri, R., Rochima, E. and Fakhrudin, S.D. (2017). Characterization of *Bacillus subtilis* and *B. licheniformis* Potentials as Probiotic Bacteria in Vanamei

Shrimp Feed (Litopenaeus Vannamei Boone, 1931). *Nusantara Bioscience*, **9**, 188-193. https://doi.org/10.13057/nusbiosci/n090214

- [78] Røssland, E., Langsrud, T., Granum, P.E. and Sørhaug, T. (2005) Production of Antimicrobial Metabolites by Strains of Lactobacillus or Lactococcus Co-Cultured with *Bacillus cereus* in Milk. *International Journal of Food Microbiology*, **98**, 193-200. https://doi.org/10.1016/j.ijfoodmicro.2004.06.003
- [79] Immink, K.A.S. and Weber, J.H. (2010) Minimum Pearson Distance Detection for Multilevel Channels with Gain and/or Offset Mismatch. *IEEE Transactions on Information Theory*, **60**, 5966-5974. <u>https://doi.org/10.1109/TIT.2014.2342744</u>
- [80] Cetin-Karaca, H. and Newman, M.C. (2018) Antimicrobial Efficacy of Phytochemicals against *Bacillus cereus* in Reconstituted Infant Rice Cereal. *Food Microbiology*, 69, 189-195. <u>https://doi.org/10.1016/j.fm.2017.08.011</u>
- [81] Guerin, A., Dargaignaratz, C., Broussolle, V., Clavel, T. and Nguyen-The, C. (2016) Combined Effect of Anaerobiosis, Low pH and Cold Temperatures on the Growth Capacities of Psychrotrophic *Bacillus cereus. Food Microbiology*, **59**, 119-123. https://doi.org/10.1016/j.fm.2016.05.015
- [82] Barbosa, J., Borges, S. and Teixeira, P. (2016) Effect of Different Conditions of Growth and Storage on the Cell Counts of Two Lactic Acid Bacteria after Spray Drying in Orange Juice. *Beverages*, 2, 8. <u>https://doi.org/10.3390/beverages2020008</u>
- [83] Fan, H., Zhang, Z.W., Li, Y., Zhang, X., Duan, Y.M. and Wang, Q. (2017) Biocontrol of Bacterial Fruit Blotch by *Bacillus subtilis* 9407 via Surfactin-Mediated Antibacterial Activity and Colonization. *Frontiers in Microbiology*, 8, 1973. <u>https://doi.org/10.3389/fmicb.2017.01973</u>
- [84] Browne, N. and Dowds, B.C. (2002) Acid Stress in the Food Pathogen Bacillus cereus. Journal of Applied Microbiology, 92, 404-414. <u>https://doi.org/10.1046/j.1365-2672.2002.01541.x</u>
- [85] Koni, T.N.I., Rusman, Hanim, C. and Zuprizal (2017) Effect of pH and Temperature on *Bacillus subtilis* FNCC 0059 Oxalate Decarboxylase Activity. *Pakistan Journal of Biological*, 20, 436-441. <u>https://doi.org/10.3923/pjbs.2017.436.441</u>
- [86] Setlow, P. (2014) Spore Resistance Properties. *Microbiology Spectrum*, 2. <u>https://doi.org/10.1128/microbiolspec.TBS-0003-2012</u>